



ANTIMICROBIAL RESISTANCE AND BIOLOGICAL PROPERTIES OF *STAPHYLOCOCCUS* SPP. ISOLATED FROM PIGS

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Cuvinte cheie: *Staphylococcus* spp., coagularea plasmei, proprietăți biologice, antibiotice, sensibilitate, rezistență.

Introduction. Representatives of the genus *Staphylococcus* spp. cause a significant proportion of diseases in animals and humans. Nowadays the problem of their acquired antibiotic resistance is an urgent concern.

Material and methods. Isolation and identification of *Staphylococcus* spp. carried out in accordance with DSTU EN 6888:2003 standard. The susceptibility of the strains to antibiotics was determined by the disk diffusion method. Interpretation of the results was carried out in accordance with the recommendations of the 8th version of EUCAST.

Results. 77 strains of *Staphylococcus* spp. Collected from sows at the farm No. 2, were isolated: hemolytic properties were detected in 90.6% (39 coagulase positive and 19 coagulase negative); 22 (56.4%) strains of coagulase-negative *Staphylococcus* spp.; 13.6% of isolates had hemolytic properties. The results of antibiograms of crops from the farm No.1: 51.8% of strains were susceptible to penicillin, 47.6% – resistant; 13.62 – susceptible to fluoroquinolones, 80.9% – resistant; 96.7% – susceptible to chloramphenicol, no resistant strains were detected.

Conclusions. Acquired resistance of *Staphylococcus* spp. to certain groups of antibiotics isolated from pigs, indicated the irrational use of antimicrobial therapy. Differences were found in the susceptibility of coagulase-positive and coagulase-negative *Staphylococcus* spp. to all groups of antibiotics.

REZISTENȚA LA ANTIMICROBIENE ȘI PROPRIETĂȚILE BIOLOGICE A *STAPHYLOCOCCUS* SPP. IZOLATE DE LA PORCINE

Introducere. Reprezentanții genului *Staphylococcus* provoacă un număr semnificativ de boli la animale și la oameni. Actualmente, o problemă majoră o prezintă rezistența dobândită a acestor tulpini la antibiotice.

Material și metode. Izolarea și identificarea *Staphylococcus* spp. s-a realizat în conformitate cu standardul DSTU EN 6888:2003. Sensibilitatea tulpinilor la antibiotice a fost determinată prin metoda disc-difuzimetrică, iar interpretarea rezultatelor a fost efectuată potrivit recomandărilor EUCAST, versiunea 8.

Rezultate. De la scroafele din ferma nr. 1 au fost izolate 77 de tulpini de *Staphylococcus* spp: proprietăți hemolitice au fost detectate la 90,6% din probe (39 coagulazo-pozitive și 19 coagulazo-negative). De la scroafele din ferma nr.2 au fost izolate 22 (56,4%) tulpini de *Staphylococcus* spp. coagulazo-negative, dintre care 13,6% posedau proprietăți hemolitice. Rezultatele antibioticogramei culturilor de la ferma nr.1 ne arată că 51,8% din culturi sunt sensibile și 47,6% sunt rezistente la peniciline; 13,6% sunt sensibile și 80,9% sunt rezistente la fluorochinolone, iar 96,7% din tulpini au fost sensibile la cloramfenicol, nefiind detectate culturi rezistente.

Concluzii. Rezistența dobândită la tulpinile de *Staphylococcus* spp, izolate de la porcinele din ferma nr.1, față de anumite grupuri de antibiotice, indică utilizarea irațională a terapiei antimicrobiene. Astfel, s-au constatat diferențe în sensibilitatea tulpinilor de *Staphylococcus* spp, coagulazo-pozitive și coagulazo-negative, la toate grupele de antibiotice.

INTRODUCTION

Staphylococcus carriers were an important source of contamination for food, raw materials, birds, pigs and pork products (1, 2). It has been repeatedly proven that pigs were sources of staphylococci, in particular methicillin-resistant *S. aureus* (MRSA) (3). Such cases were especially common in Denmark (4), Canada (4), Germany (6) and Switzerland (7). In young pigs, the disease manifests itself in the form of exudative epidermitis (EE), caused by strains of *Staphylococcus hyicus*, *Staphylococcus aureus* and *Staphylococcus chromogenes*, which produce exfoliative toxins. However, pigs in most cases were hidden carriers (7). Experimental transfer of methicillin-resistant *S. aureus* to minks during feeding of pork waste contaminated with MRSA is also known (9). In 2017, 80 methicillin-resistant staphylococcus strains were isolated in Ukraine, including 77.5% from domestic animals, 11.3% from poultry, 6.3% from cattle and 5% from pigs (10).

The purpose of the study was to investigate the biological properties and antibiotic resistance of *Staphylococcus* spp. isolated from pigs of two industrial pig farms located in Kyiv (No.1) and Vinnytsya regions (No.2).

MATERIAL AND METHODS

Isolation and identification of *Staphylococcus* spp. was conducted in accordance with: DSTU EN 6888:2003 "Microbiology of food and animal feed" standard was cultivated to the Baird Parker agar (Merck) medium and were incubated at 37°C for 24-48 hours. Isolates produced on Baird Parker agar at the end of this period were defined by morphology colony, Gram stain, catalase test, and coagulation test. The hemolytic properties of staphylococci were studied on Columbia blood agar (BioMerieux). Selected colonies from Columbia blood agar introduced into Tryptone-soy broth and cultured at t 35°C for 2-4 hours. The optical density was determined using a densitometer DEN-1(Biosan) and McFarland standard (HiMedia). The susceptibility of strains to antibiotics was determined by disc-diffusion method using Müller-Hinton agar, inoculum: 0.5 according to the McFarland turbidity standard and were incubated at 35°C, for 18±2 hours. The results were assessed in accordance with the recommendations of version 8 of the European Committee on Antimicrobial Susceptibility Testing EUCAST (11)

and Guidelines "Determination of susceptibility of microorganisms to antibacterial drugs" (12).

RESULTS

Farm No. 1 proved to be a satisfactory farm in terms of safety against contagious diseases; number of pigs was up to 2 500 heads. There were 210 pigs. Purulent diseases, abscesses and boils were not reported in piglets. Of the 77 sows sampled from the nose *Staphylococcus* spp. were isolated in 64 cases (83.1%). All strains grown on the Beard Parker agar had typical colonies of black and grey, shiny and convex with a diameter of 1 mm to 1.5 mm after incubation for 24 hours and a diameter of 1.5-2.5 mm after 48 hours of incubation. The 39 isolates (60.94%) were coagulated with rabbit plasma. The ability to hemo-lysis of sheep erythrocytes showed only 58 (90.6%) of strains. Of these, 39 coagulase-positive strains and 19 coagulase-negative strains did not show hemolytic properties of 6 coagulase-negative strains.

The farm No. 2 proved to be a satisfactory farm in terms of contagious diseases; number of pigs was up to 3 500 heads. Diseases caused by *Staphylococcus* spp. we're not registered. In 22 samples (56.4%), isolates of *Staphylococcus* spp. were isolated from 39 sows from farm No.2.

The isolates grew in the form of shiny black and grey colonies with a narrow white margin. The isolates did not coagulate plasma. The 3 strains (13.6%) from the isolated strains had hemolytic properties.

Staphylococcus had a natural susceptibility to penicillins, but subsequently acquired resistance to them (13). In the study, were used: Natural Penicillin, Benzylpenicillin; Semisynthetic Oxacillin, Gentamicin, Tobramycin from the aminoglycosides group, and Erythromycin from the macrolids group (13). The susceptibility to natural Tetracycline and Semisynthetic Doxycycline; Lincomycin and Clindamycin from the group of lincosamides was investigated. Susceptibility for norfloxacin was recommended for screening susceptibility for all fluoroquinolones (11). There were studied Norfloxacin, Ciprofloxacin, Ofloxacin, Pefloxacin, Lomefloxacin, Levofloxacin, Sparfloxacin and Gatifloxacin (12, 13). Susceptibility to Chloramphenicol, and Rifampicin was investigated (11).

51.87% of the selected strains obtained at the farm No.1 were susceptible to Penicillin antibiotics,

and 47.59% were resistant. 78.48% of bacterial strains showed susceptibility to aminoglycosides, and 21.52% of the strains were resistant. 87.1% of strains showed susceptibility to macrolides. 70.45% of the isolates were susceptible to tetracyclines. 70% of isolates showed susceptibility, and 30% were resistant to lincosamides. 13.62% were susceptible, and 80.93% – resistant to fluoroquinolones. 96.72% of revealed strains were susceptible to chloramphenicol. 73.53% of the studied isolates were susceptible to rifampicin.

10 strains from 22 pigs, on farm No. 2, were isolated and an antibiogram was determined (tab. 1). It was established that 10 isolated strains were resistant to oxacillin and clindamycin. 3 strains were susceptible, and 7 – resistant to benzylpenicillin and norfloxacin. 1 culture was susceptible, and 9 – resistant to tetracycline and erythromycin. 4 strains were susceptible, and 6 – resistant to chloramphenicol.

Table 1. Ranges of growth inhibition diameters of the studied cultures isolated from pigs on farm No. 2, mm. Min-Max.

Name of antibiotic	Eucast interpretation (version 8.0) and Guidelines “Determination of susceptibility of microorganisms to antibacterial drugs”	Ranges of diameters of cultures growth inhibition mm. Min-Max	S strains (n)	R strains (n)
Benzylpenicillin	26>s; 26<r	0-29	3	7
Oxacillin	18≥s; 17<r	0-16	0	10
Erythromycin	21>s; 18<r	16-25	1	9
Clindamycin	22>s; 19<r	0-17	0	10
Norfloxacin	24>s; 24<r	0-28	3	7
Tetracycline	22>s; 19<r	17-24	1	9
Chloramphenicol	18>s; 18<r	15-22	4	6

Note: “0” – continuous growth, “15-22” – the minimum and maximum value of growth inhibition of the test culture

Number of coagulase-positive and coagulase-negative strains isolated from the farm No. 1 was showed in Table 2.

There was 100% resistance to benzylpenicillin among 18 coagulase-positive strains of *Staphylococcus* spp. from isolated strains. Oxacillin: intermediate resistance (12) – 3% of isolated strains, resistant – 6%, susceptible – 91% of strains. All resistant strains. Doxycycline: 28.5% – intermediate resistance; 28.5% – resistant and 43% susceptible strains.

There were 7.8% of the strains from which 92% were resistant to lincomycin. Clindamycin: 42% – resistant, 58% of susceptible isolates. 100% of the coagulase-positive *Staphylococcus* spp. were resistant to Norfloxacin and Ciprofloxacinum. 4.7% and 95.3% of resistant strains were susceptible to Ofloxacin. Pefloxacin: 10% – resistant, 60% – intermediate resistance and 30% susceptible isolates. Lomefloxacin was found to be resistant in 87% of strains, intermediate in 6.5% and susceptible in 6.5% of strains. 6.7% of the isolated strains were susceptible to Levofloxacin and the remaining 93.3% were resistant There were 97%

cultures resistant and 3% isolates which showed susceptibility to Sparfloxacin. Gatifloxacin: intermediate resistance strains – 13% and 87% – resistant strains. The 84.6% demonstrated resistance to rifampicin 7.7% and intermediate resistance 7.7% of strains.

In the group of coagulase-negative *Staphylococcus* spp. isolated from pigs in farm No. 1, there were 25% resistant strains and 75% strains showed susceptibility to Benzylpenicillin. All 100% strains were susceptible to Oxacillin. Ampicillin: 13% of strains were susceptible, 87% of strains – resistant. Gentamicin: 46% of strains were susceptible and 54% – resistant. There were 92% susceptible and 8% resistant strains to Tobramycin. Erythromycin: 6.9% of strains were resistant and 6.9%-intermediate resistance, the rest all isolated strains (86.2%) showed susceptibility.

Tetracycline: 81.5% of strains were intermediate resistance, 11% – resistant, and 7.5% strains were susceptible. Doxycycline: 4% of strains were intermediate resistance, 4% – resistant, and 92% susceptible isolates. Lincomycin: 84% of the strains were susceptible and 16% were resistant.

3% intermediate resistance strains to Clindamycin, the remaining 97% – susceptible. Norfloxacin: susceptible – 3% and intermediate resistance – 3% and 94% – resistant strains. 80% of strains were 20% susceptible to Ciprofloxacin. Ofloxacin: susceptible – 25%, resistant – 75% of isolates. Pefloxacin: resistant – 50%, intermediate resistance – 23%, susceptible – 27% of strains.

Lomefloxacin: susceptible – 16%, intermediate resistance – 8%, and resistant – 76% isolates. 8% of

the strains were susceptible to Levofloxacin and 92% were resistant. 64.5% of the strains showed susceptibility to Sparfloxacin and 35.5% was resistant. There were 23% intermediate resistance and 8% resistant and 69% of strains susceptible to Gatifloxacin. Chloramphenicol: intermediate resistance – 7% of strains from 93% of strains. Rifampicin: 66.5% were susceptible, 24% were intermediate resistance, and 9.5% of isolates were resistance.

Table 2. Number of studied cultures of *Staphylococcus* spp. isolated in the farm No.1.

Name of antibiotic	The number of cultures of <i>Staphylococcus</i> spp.							
	Coagulase-positive				Coagulase-negative			
	S	I	R	Total	S	I	R	Total
Penicillins								
Benzyloxyethylpenicillin	0	0	18	18	7	0	21	28
Oxacillin	30	1	2	33	31	0	0	31
Ampicillin	0	0	14	14	3	0	20	23
Aminoglycosides								
Gentamicin (I)	13	0	1	14	12	0	14	26
Tobromycin (II)	14	0	0	14	23	0	2	25
Macrolides								
Erythromycin	29	2	2	33	25	2	2	29
Tetracyclines								
Tetracycline	10	0	11	21	22	3	2	27
Doxycycline	6	4	4	14	24	1	1	26
Linkosamides								
Lincomycin	1	0	12	13	21	0	4	25
Clindamycin	18	0	13	31	30	1	0	31
Fluoroquinolones								
Norfloxacin (II)	0	0	27	27	1	1	28	30
Ciprofloxacin (II)	0	0	15	15	5	0	20	25
Ofloxacin (II)	1	0	20	21	7	0	21	28
Pefloxacin (II)	3	6	1	10	7	6	13	26
Lomefloxacin (II)	1	1	13	15	4	2	19	25
Levofloxacin (III)	1	0	14	15	2	0	23	25
Sparfloxacin (III)	1	0	32	33	11	0	20	31
Gatifloxacin (IV)	0	2	13	15	6	2	18	26
Others								
Chloramphenicol	32	0	0	32	27	2	0	29
Rifampicin	11	1	1	13	14	5	2	21

DISCUSSIONS

The obtained results indicate a significant colonization of coagulase-positive and coagulase-negative staphylococci in the nasal passages of sows. A significant percentage of isolated crops possesses “pathogenicity factors”. Plasma coagulation and hemolysis were particularly important. The high circulation of staphylococci in the herd leads to an

increase of antibiotic resistance. Natural penicillin is known to have little effect on staphylococci, while synthetic and semisynthetic penicillin antibiotics of other groups inhibit their growth.

The results showed that in the farm No. 1 polyresistant strain of coagulase-positive and coagulase-negative staphylococci was present. A major

component of beta-lactam resistance is the so-called *mecA* gene, which encodes the formation of modified penicillin-binding protein and thus interferes with the incorporation of beta-lactam into the cell wall. When the cell is methicillin-resistant *Staphylococcus aureus* in contact with β -lactam antibiotics, the additional β -lactam-resistant penicillin binding protein (PBP2a) takes on the biosynthetic functions of normal PBPs.

Staphylococcus aureus resistance to methicillin (Oxacillin) may be due to the production of additional PBP-2a, which is encoded by the chromosomal *mecA* gene, inactivation through hyperproduction of β -lactamases and modification of normal PBPs. The presence two resistant strains of coagulase-positive staphylococci and one intermediate resistance in pigs may indicate the possible presence of the *mecA* gene. Also, the resistance of the isolated strains to Ampicillin indicates the synthesis of staphylococcal penicillinase. The higher resistance to Tetracycline was shown by coagulase-positive staphylococci, which was

confirmed by the resistance of these strains to Doxycycline. Also, indicative is the resistance of some coagulase-positive staphylococci to both Lycomycin and Clindamycin, which is not observed among coagulase-negative strains. Practically all staphylococci have been shown to be resistant to Fluoroquinolones of different generations, including Gatifloxacin. Two intermediate resistance strains of coagulase-negative staphylococci were detected for Chloramphenicol while all coagulase-positive strains were susceptible.

As a result, the circulation of polyresistant coagulase-positive staphylococcus strains proved to be resistant to almost all antibiotic groups, especially to Fluoroquinolones – 4.6% susceptible, 6.0% – intermediate resistance, and 89.4% – resistant. Among coagulase-negative staphylococci, 20% were susceptible, 5.0% were intermediate resistance, and 75% were resistant to Fluoroquinolones. Coagulase-negative strains from farm No. 2 also had poly resistant properties in tested antibiotics of all groups.

CONCLUSIONS

1. High percentage of staphylococcus circulation in pigs of experimental farms was detected.
2. The selected strains possessed “pathogenicity factors” by hemolysis and plasma coagulation.
3. Some staphylococci showed resistance to three or more antibiotics at the same time.
4. All these factors testify the misuse of antibiotics and the rapid manifestation of resistance to their individual representatives.
5. To confirm the emergence of the mechanism of resistance, it is necessary to carry out molecular genetic studies of isolated strains.
6. It is necessary to change fundamentally the pattern of antibiotic use in pigs in order to prevent staphylococcus resistance and their subsequent transfer to humans.

CONFLICT OF INTERESTS

All authors declare no competing interests.

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